

Part Two: BRGSF, a New Immunodeficient Model for Immuno-Oncology Studies

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As discussed in our previous commentary, mice are the experimental models of election in preclinical research, as they represent an affordable, rapidly reproducing and easily maintained mammalian model to study disease etiology and therapeutic assessment.^{1,2} Syngeneic tumor transplantation and genetically engineered mouse models have offered tremendous insight in the basic mechanisms of immune regulation. However, for many years, substantial genetic and genomic differences between humans and mice have hampered the translatability of experimental findings in infectious and immune-related diseases, very often leading to clinical trial failures.^{3,4}

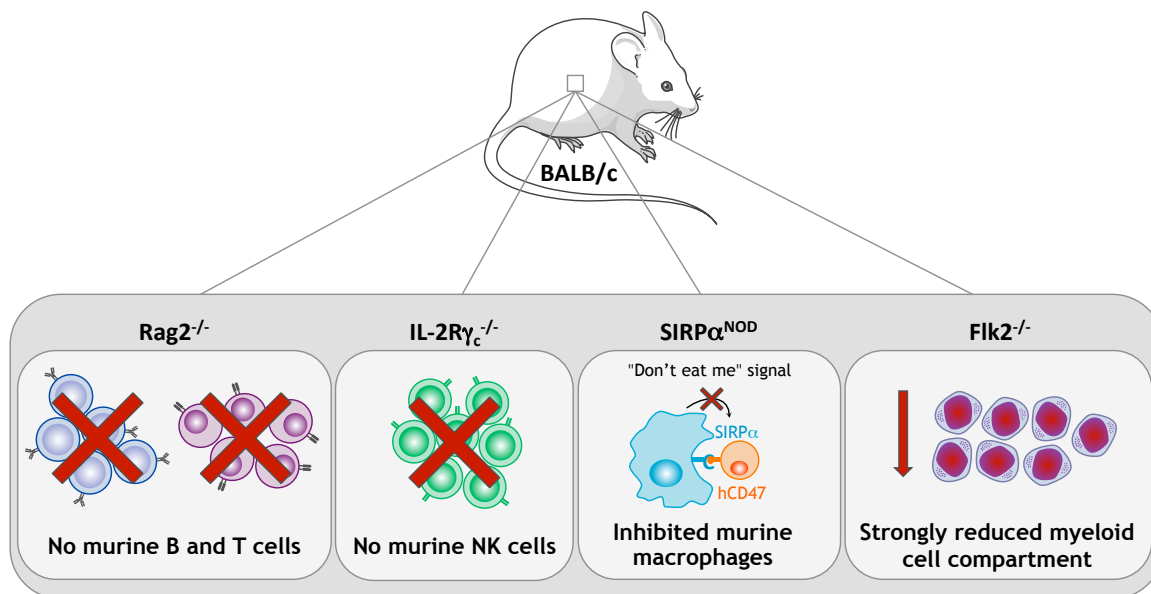
In order to bridge this gap and enhance the transfer of knowledge from *in vitro* and mouse-based investigations toward clinical applications, scientists have generated a wide variety of immunodeficient models such as SCID, NUDE and Rag-deficient mice engrafted either subcutaneously or orthotopically with cell-derived xenografts (CDXs) or patient-derived xenografts (PDXs).^{5,6}

NUDE mice were mainly used to investigate cancer initiation and progression, as these animals could be easily engrafted subcutaneously due to the absence of fur. SCID and Rag-deficient mice, on the other hand, were the perfect hosts to study blood cancers such as lymphoma, myeloma, and leukemia, as they are more immunodeficient (i.e., they both lack T and B cells) than NUDE mice.⁵ All these models, however, still possessed residual murine immune cells and, therefore, could not be used for long-term xenograft studies. Moreover, the lack of functional immune system was of major concern for a proper understanding of the pathophysiology of tumor development, escape mechanism to immune system, and assessment of immunotherapies enabling the development of effective anti-tumor response.^{2,7}

For all these reasons, scientists developed a new generation of highly immunodeficient mice by backcrossing SCID and Rag-deficient mice into non-obese diabetic (NOD) or Balb/c mice. The resulting strains, NOG (NOD/Shi-scid IL-2 γ null), NSG (NOD-scid IL-2 γ ^{-/-}) and BRGS (Balb/c Rag^{2tm1Fwa} IL-2 γ ^{tm1Cgn} Sirp α ^{NOD}), carry multiple genetic defects, including mutations in i) IL-2 receptor common γ -chain (IL-2 γ), leading to profound NK cells deficiency, ii) Prkdc (NOG and NSG) or Rag2 (BRGS), causing a depletion in B and T cells, and iii) NOD-specific polymorphic Sirp α , resulting in a reduced phagocytosis of human CD47⁺ cells by murine macrophages (i.e., 'don't eat me' signal).⁸⁻¹¹

The NOG and NSG were largely used in PDX studies, as they sustain better engraftments of human tissues compared to NUDE and SCID mice. However, due to the mutations in Prkdc, these strains show the SCID side effect of high sensitivity to radiation, T-cell leakage, and increased incidence of thymic lymphoma formation; as such, they cannot be used to predict clinical response to certain anticancer drugs, or for long-term transplantation studies.^{12,13} The BRGS solves both these problems because it does not carry the Prkdc mutation and, therefore, it does not possess the SCID phenotype observed in the NSG and NOG strain.^{9,10}

The BRGS has been recently improved by a team of scientists at the Pasteur Institute, by backcrossing it into the BRGF (BALB/c Rag^{2tm1Fwa} Il2r^{tm1Cgn} Flt3^{tm1lr1}) strain. The resultant BRGSF (BALB/c Rag^{2tm1Fwa} Il2r^{tm1Cgn} Sirp^{αNOD} Flt3^{tm1lr1}) is highly immunodeficient, because besides carrying the genetic defects of its parental strains, it possesses a deficiency in the fetal liver kinase-2 (Flk2), which regulates the development of the myeloid compartment.^{14–16}



For all these reasons, the BRGSF mouse model represents a valuable tool for vaccine development, efficacy and safety of chimeric antigen receptor (CAR) T cell therapy, and myeloid compartment development studies. Moreover, as it is highly permissive to human hematopoietic cells engraftment, BRGSF serves as the optimal model to generate human immune system (HIS) mice to study and predict human immune responses *in vivo*.^{8,14–16}

In part three, we will discuss HIS mouse models in more detail.

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